: .

O S L13 AND (PH OR PI) AND DALTON#

3 S L20 AND PI

0 S L13 AND DALTON#

11 S L13 AND WEIGHT 9 S L24 AND (PH OR PI)

L21

L22

L23 L24

L25

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SO

L25 ANSWER 1 OF 9 HCAPLUS COPYRIGHT 2001 ACS ACCESSION NUMBER: 1998:676139 HCAPLUS DOCUMENT NUMBER: 130:11851 Purification and characterization of trehalose TITLE: phosphorylase from the commercial mushroom Agaricus bisporus Wannet, Wim J. B.; Huub, J. M.; Den Camp, Op; AUTHOR(S): Wisselink, Hendrik W.; Van Der Drift, Chris; Van Griensven, Leo J. L. D.; Vogels, Godfried D. Department of Microbiology, Faculty of Science, CORPORATE SOURCE: University of Nijmegen, Nijmegen, NL-6525, Neth. Biochim. Biophys. Acta (1998), 1425(1), SOURCE: 177-188 CODEN: BBACAQ; ISSN: 0006-3002 Elsevier Science B.V. PUBLISHER: Journal DOCUMENT TYPE: English LANGUAGE: Trehalose phosphorylase (EC 2.4.1.64) (I) from A. AB bisporus was purified for the 1st time from a fungus. I appears to play key role in trehalose metab. in A. bisporus since no trehalase or trehalose synthase activities could be detected in this fungus. I catalyzes the reversible reaction of degrdn. (phosphorolysis) and synthesis of trehalose. Native I was found to have a mol. wt. of 240 kDa and to consist of 4 identical 61-kDa subunits. The pI of I was 4.8. The optimum temp. for both enzyme reactions was 30.degree.. The optimum pH ranges for trehalose degrdn. and synthesis were 6.0-7.5 and 6.0-7.0, resp. Trehalose degrdn. was inhibited by ATP and trehalose analogs, whereas the synthetic activity was inhibited by inorg. phosphate (Pi; Ki = 2.0 mM). I was highly specific for trehalose, Pi, glucose, and .alpha.-glucose 1-phosphate. stoichiometry of the reaction between trehalose, Pi, glucose, and .alpha.-glucose 1-phosphate was 1:1:1:1 (molar ratio). The Km values were 61, 4.7, 24 and 6.3 mM for trehalose, Pi, glucose, and .alpha.-glucose 1-phosphate, resp. Under physiol. conditions, A. bisporus I probably performs both synthesis and degrdn. of trehalose. REFERENCE COUNT: (2) Baars, J; Microbiology 1994, V140, P1161 HCAPLUS REFERENCE(S): (3) Bartlett, G; J Biol Chem 1959, V234, P466 HCAPLUS (4) Becker, A; Experientia 1996, V52, P433 HCAPLUS (5) Belocopitow, E; Biochim Biophys Acta 1970, V198, P151 HCAPLUS (6) Bergmeyer, H; Methoden der Enzymatischen Analyse 1974, P1250 HCAPLUS ALL CITATIONS AVAILABLE IN THE RE FORMAT Purification and characterization of trehalose ΤI phosphorylase from the commercial mushroom Agaricus bisporus

CODEN: BBACAQ; ISSN: 0006-3002

AB Trehalose phosphorylase (EC 2.4.1.64) (I) from A. bisporus was purified for the 1st time from a fungus. I appears to play

key role in trehalose metab. in A. bisporus since no trehalase or trehalose synthase activities could be detected in this fungus. I catalyzes the reversible reaction of degrdn. (phosphorolysis) and synthesis of trehalose. Native I was found to have a mol. wt.

Biochim. Biophys. Acta (1998), 1425(1), 177-188

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of 240 kDa and to consist of 4 identical 61-kDa subunits.
    of I was 4.8. The optimum temp. for both enzyme reactions was
30.degree..
    The optimum pH ranges for trehalose degrdn. and synthesis were
    6.0-7.5 and 6.0-7.0, resp. Trehalose degrdn. was inhibited by ATP and
    trehalose analogs, whereas the synthetic activity was inhibited by inorg.
    phosphate (Pi; Ki = 2.0 \text{ mM}). I was highly specific for
    trehalose, Pi, glucose, and .alpha.-glucose 1-phosphate.
    stoichiometry of the reaction between trehalose, Pi, glucose,
    and .alpha.-glucose 1-phosphate was 1:1:1:1 (molar ratio). The Km values
    were 61, 4.7, 24 and 6.3 mM for trehalose, Pi, glucose, and
     .alpha.-glucose 1-phosphate, resp. Under physiol. conditions, A.
bisporus
     I probably performs both synthesis and degrdn. of trehalose.
     trehalose phosphorylase mushroom; Agaricus
ST
     trehalose phosphorylase
    Michaelis constant
ΙT
        (of trehalose phosphorylase from the com. mushroom
        Agaricus bisporus)
     Agaricus bisporus
ΙT
        (purifn. and characterization of trehalose
        phosphorylase from the com. mushroom Agaricus bisporus)
     37205-59-7P, Trehalose phosphorylase
IΤ
    RL: BAC (Biological activity or effector, except adverse); PRP
     (Properties); PUR (Purification or recovery); BIOL (Biological study);
     PREP (Preparation)
        (purifn. and characterization of trehalose
        phosphorylase from the com. mushroom Agaricus bisporus)
L25 ANSWER 2 OF 9 HCAPLUS COPYRIGHT 2001 ACS
                         1998:314518 HCAPLUS
ACCESSION NUMBER:
                         129:25076
DOCUMENT NUMBER:
TITLE:
                         A thermostable trehalose
                         phosphorylase of Thermoanaerobium and its uses
                         in the preparation of glucosides
                         Nakada, Tetsuya; Kubota, Michio; Chaen, Hiroto;
INVENTOR(S):
                         Miyake, Toshio
                         Kabushiki Kaisha Hayashibara Seibutsu Kagaku
PATENT ASSIGNEE(S):
Kenkyujo,
                         Japan
                         Eur. Pat. Appl., 39 pp.
SOURCE:
                         CODEN: EPXXDW
DOCUMENT TYPE:
                         Patent
                         English
LANGUAGE:
FAMILY ACC. NUM. COUNT:
PATENT INFORMATION:
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KIND DATE	E AP	PLICATION NO.	DATE
A2 1998	30513 EP	1997-308980	19971107 <
A3 1999	90721		
E, CH, DE, DK,	, ES, FR, GB,	GR, IT, LI, LU	, NL, SE, MC, PT,
•			
A2 1998	31117 JP	1997-319139	19971106
A 1998	31201 US	1997-966389	19971107
A 1999	90608 US	1998-102644	19980623
A 1999	90302 US	1998-103509	19980624
A 1999	91130 US	1998-218032	19981222
· · · · · · · · · · · · · · · · · · ·	JP 19	96-311232	19961108
	JP 19	97-61716	19970303
	A2 1998 A3 1999 E, CH, DE, DK, I A2 1998 A 1999 A 1999 A 1999	A2 19980513 EP A3 19990721 E, CH, DE, DK, ES, FR, GB,  A2 19981117 JP A 19981201 US A 19990608 US A 19990302 US A 19991130 US FO.: JP 19	A2 19980513 EP 1997-308980 A3 19990721 E, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU,  A2 19981117 JP 1997-319139 A 19981201 US 1997-966389 A 19990608 US 1998-102644 A 19990302 US 1998-103509 A 19991130 US 1998-218032

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US 1997-966389
                    19971107
US 1998-103509
                    19980624
```

A thermostable trehalose phosphorylase is obtained AB from microorganisms of the genus Thermoanearobium that hydrolyzes trehalose in the presence of an inorg. phosphoric acid to form D-glucose and .beta.-D-glucose-1-phosphate is described. The enzyme can use .beta.-D-glucose-1-phosphate as a saccharide donor to create novel glucosides such as glucosyl-D-galactoside, that are known but rare and they can be produced on an industrial-scale and at a relatively-low cost. The enzyme has a mol. wt. of 88,000, an isoelec. point of 5.4.+-.0.5, a temp. optimum of 70.degree., a pH optimum of 7.0-7.5, is activated by thiols and inhibited by divalent cations. stable at 60.degree. for an hour. The protein may be manufd. by expression of the cloned gene. ΤI

A thermostable trehalose phosphorylase of Thermoanaerobium and its uses in the preparation of glucosides

EP 841397 A2 19980513 PΤ KIND DATE APPLICATION NO. DATE PATENT NO. 19971107 <--EP 841397 A2 19980513 EP 1997-308980 PΤ A3 19990721 EP 841397 R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI JP 1997-319139 19971106 A2 19981117 JP 10304881 19981201 US 1997-966389 19971107 US 5843748 Α US 5910436 Α 19990608 US 1998-102644 19980623 19990302 US 1998-103509 19980624 US 5876975 Α 19991130 US 1998-218032 19981222 US 5993889 Α

A thermostable trehalose phosphorylase is obtained from microorganisms of the genus Thermoanearobium that hydrolyzes trehalose in the presence of an inorg. phosphoric acid to form D-glucose and .beta.-D-glucose-1-phosphate is described. The enzyme can use .beta.-D-glucose-1-phosphate as a saccharide donor to create novel glucosides such as glucosyl-D-galactoside, that are known but rare and they can be produced on an industrial-scale and at a relatively-low cost. The enzyme has a mol. wt. of 88,000, an isoelec. point of 5.4.+-.0.5, a temp. optimum of 70.degree., a **pH** optimum of 7.0-7.5, is activated by thiols and inhibited by divalent cations. stable at 60.degree. for an hour. The protein may be manufd. by expression of the cloned gene.

thermostable trehalose phosphorylase Thermoanaerobium STgene cloning; sweetener prepn thermostable trehalose phosphorylase Thermoanaerobium; glucoside prepn thermostable trehalose phosphorylase Thermoanaerobium

ΙT Radish (Raphanus sativus)

(enzymic prepn. of glucosides for use as sweeteners for pickling of; thermostable trehalose phosphorylase of

Thermoanaerobium and its uses in prepn. of glucosides)

IΤ Candy

AΒ

Chewing gum Chocolate

Dentifrices

Desserts

Milk preparations

Pickles

(enzymic prepn. of glucosides for use as sweeteners for; thermostable trehalose phosphorylase of Thermoanaerobium and its uses in prepn. of glucosides)

IΤ Sweetening agents

(enzymic prepn. of glucosides for use as; thermostable

```
trehalose phosphorylase of Thermoanaerobium and its
        uses in prepn. of glucosides)
ΙT
     Skin creams
        (enzymic prepn. of glucosides for use in; thermostable
        trehalose phosphorylase of Thermoanaerobium and its
        uses in prepn. of glucosides)
ΙT
     Nutrients
        (for intubation, enzymic prepn. of glucosides for use as sweeteners
        for; thermostable trehalose phosphorylase of
        Thermoanaerobium and its uses in prepn. of glucosides)
ΙT
     Genes (microbial)
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (for trehalose pyrophosphorylase of Thermoanaerobium brockii, cloning
        and expression of; thermostable trehalose
        phosphorylase of Thermoanaerobium and its uses in prepn. of
        glucosides)
     DNA sequences
TT
        (for trehalose pyrophosphorylase of Thermoanaerobium brockii;
        thermostable trehalose; phosphorylase of
        Thermoanaerobium and its uses in prepn. of glucosides)
     Protein sequences
IT
        (of trehalose pyrophosphorylase of Thermoanaerobium brockii;
        thermostable trehalose phosphorylase of
        Thermoanaerobium and its uses in prepn. of glucosides)
ΙT
     Jams and Jellies
        (strawberry, enzymic prepn. of glucosides for use as sweeteners for;
        thermostable trehalose phosphorylase of
        Thermoanaerobium and its uses in prepn. of glucosides)
     Milk preparations
TT
        (sweetened condensed milk, enzymic prepn. of glucosides for use as
        sweeteners for; thermostable trehalose phosphorylase
        of Thermoanaerobium and its uses in prepn. of glucosides)
     Thermoanaerobacter brockii brockii
ΙT
     Thermoanaerobium
        (thermostable trehalose phosphorylase of
        Thermoanaerobium and its uses in prepn. of glucosides)
TT
     Glycosides
     RL: BMF (Bioindustrial manufacture); BPN (Biosynthetic preparation); BIOL
     (Biological study); PREP (Preparation)
        (trehalose pyrophosphorylase for manuf. of; thermostable
        trehalose phosphorylase of Thermoanaerobium and its
        uses in prepn. of glucosides)
                   208064-41-9
     208064-39-5
IT
     RL: BOC (Biological occurrence); BSU (Biological study, unclassified);
CAT
     (Catalyst use); PRP (Properties); BIOL (Biological study); OCCU
     (Occurrence); USES (Uses)
        (amino acid sequence; thermostable trehalose
        phosphorylase of Thermoanaerobium and its uses in prepn. of
        glucosides)
     14048-34-1, .beta.-D-Glucose-1-phosphate
TΤ
     RL: BPR (Biological process); RCT (Reactant); BIOL (Biological study);
     PROC (Process)
        (as substrate of trehalose pyrophosphorylase; thermostable
        trehalose phosphorylase of Thermoanaerobium and its
        uses in prepn. of glucosides)
                                           207570-55-6P
                           207570-54-5P
                                                          208041-60-5P
     99-20-7P, Trehalose
ΙT
     208041-61-6P
     RL: BPN (Biosynthetic preparation); BIOL (Biological study); PREP
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(Preparation)
        (enzymic prepn. of; thermostable trehalose
       phosphorylase of Thermoanaerobium and its uses in prepn. of
        glucosides)
     50-99-7, D-Glucose, reactions
                                     58-86-6, D-Xylose, reactions
                                                                    59-23-4,
ΙT
                                                        2438-80-4, L-Fucose
    D-Galactose, reactions
                              154-17-6, 2-Deoxyglucose
                              3458-28-4, D-Mannose 3615-37-0, D-Fucose
    3416-24-8, Glucosamine
    7512-17-6, N-Acetyl glucosamine
    RL: RCT (Reactant)
        (glucosidation of; thermostable trehalose
        phosphorylase of Thermoanaerobium and its uses in prepn. of
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     208064-40-8
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    RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (nucleotide sequence; thermostable trehalose
        phosphorylase of Thermoanaerobium and its uses in prepn. of
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                   207570-57-8
                                 207570-58-9
IT
     207570-56-7
     RL: BSU (Biological study, unclassified); PRP (Properties); BIOL
     (Biological study)
        (peptide of trehalose phosphorylase of
        Thermoanaerobium; thermostable trehalose
        phosphorylase of Thermoanaerobium and its uses in prepn. of
        glucosides)
     37205-59-7, Trehalose phosphorylase
ΙT
     RL: BOC (Biological occurrence); BSU (Biological study, unclassified);
CAT
     (Catalyst use); PRP (Properties); BIOL (Biological study); OCCU
     (Occurrence); USES (Uses)
        (thermostable trehalose, phosphorylase of
        Thermoanaerobium and its uses in prepn. of glucosides)
L25 ANSWER 3 OF 9 HCAPLUS COPYRIGHT 2001 ACS
                         1997:579816 HCAPLUS
ACCESSION NUMBER:
                         127:187873
DOCUMENT NUMBER:
                         Enzymic method for determining 1,5-anhydroglucitol
TITLE:
for
                         diagnosis of diabetes
                         Aisaka, Kazuo; Tazoe, Sakae; Ando, Katsuhiko; Ochiai,
INVENTOR(S):
                         Keiko
                         Kyowa Hakko Kogyo Co., Ltd., Japan; Aisaka, Kazuo;
PATENT ASSIGNEE(S):
                         Tazoe, Sakae; Ando, Katsuhiko; Ochiai, Keiko
                         PCT Int. Appl., 55 pp.
SOURCE:
                         CODEN: PIXXD2
DOCUMENT TYPE:
                         Patent
                         Japanese
LANGUAGE:
FAMILY ACC. NUM. COUNT:
PATENT INFORMATION:
                      KIND
                            DATE
                                           APPLICATION NO.
     PATENT NO.
                            _____
                            19970828
                                           WO 1997-JP440
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                      A1
     WO 9731103
         W: AU, BG, BR, CA, CN, CZ, HU, JP, KR, MX, NO, NZ, PL, RO, SG, SI,
             SK, UA, US, VN, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
         RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT,
SE
                                           CA 1997-2218488
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                       AΑ
                            19970828
     CA 2218488
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     AU 9717327
                       B2
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19980225
                                      EP 1997-904571 19970219 <--
                      A1
    EP 825258
        R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
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                           19980429
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    US 6153419
                      Α
                           20001128
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                                       JP 1996-32393
PRIORITY APPLN. INFO .:
                                       WO 1997-JP440
                                                        W
                                                           19970219
    A method for detg. 1,5-anhydroglucitol (I) by using an enzyme that is
    susceptible to the inhibition by I in a concn.-dependent manner is
    disclosed. The method employs a compn. consisting of the enzyme (e.g.
    trehalase and trehalose phosphorylase), its substrate,
    and a reagent for the detn. of the enzymic reaction product.
    disclosed are a novel trehalase prepd. from Nocardia, exhibiting a
    pH optimum 5-6, temp. optimum 45.degree., Km <0.33 mM I or 6.7 mM
    trehalose, and mol. wt. 90,000 by SDS-PAGE or 400,000 by gel
    filtration. The method is useful for the diagnosis of diabetes. A few
    compn. and a blood anal. were demonstrated.
    WO 9731103 A1 19970828
PΙ
                     KIND DATE
                                          APPLICATION NO. DATE
    PATENT NO.
                     A1 19970828
                                         WO 1997-JP440 19970219 <--
PΙ
    WO 9731103
        W: AU, BG, BR, CA, CN, CZ, HU, JP, KR, MX, NO, NZ, PL, RO, SG, SI,
            SK, UA, US, VN, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
        RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT,
SE
                                          CA 1997-2218488 19970219 <--
                           19970828
    CA 2218488
                      AA
                                          AU 1997-17327
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    AU 9717327
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                           20000810
                      B2
    AU 722636
                                          EP 1997-904571
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                      A1
                           19980225
    EP 825258
        R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
            IE, FI
                                          CN 1997-190096
                                                           19970219 <--
                           19980429
                      Α
    CN 1180378
                           20001128
                                          US 1997-930709
                                                           19971016
    US 6153419
                      Α
    A method for detg. 1,5-anhydroglucitol (I) by using an enzyme that is
    susceptible to the inhibition by I in a concn.-dependent manner is
    disclosed. The method employs a compn. consisting of the enzyme (e.g.
    trehalase and trehalose phosphorylase), its substrate,
    and a reagent for the detn. of the enzymic reaction product. Also
    disclosed are a novel trehalase prepd. from Nocardia, exhibiting a
    pH optimum 5-6, temp. optimum 45.degree., Km <0.33 mM I or 6.7 mM
    trehalose, and mol. wt. 90,000 by SDS-PAGE or 400,000 by gel
    filtration. The method is useful for the diagnosis of diabetes. A few
    compn. and a blood anal. were demonstrated.
     Catellatospora ferruginea
ΙT
        (trehalose phosphorylase prepd. from; enzymic
       method for detg. 1,5-anhydroglucitol for diagnosis of diabetes)
ΙT
     9025-52-9, Trehalase 37205-59-7, Trehalose
    phosphorylase
    RL: ARG (Analytical reagent use); BAC (Biological activity or effector,
    except adverse); ANST (Analytical study); BIOL (Biological study); USES
     (Uses)
        (reagent compn. contg.; enzymic method for detg. 1,5-anhydroglucitol
        for diagnosis of diabetes)
L25 ANSWER 4 OF 9 HCAPLUS COPYRIGHT 2001 ACS
                         1997:187051 HCAPLUS
ACCESSION NUMBER:
                         126:183170
DOCUMENT NUMBER:
                         Preparation, thermostability, and synthetic use of
TITLE:
                        heat-resistant maltose phosphorylase from Bacillus
                         Ishii, Keiko; Inoue, Yasushi; Tomita, Tetsuji
INVENTOR(S):
```

PATENT ASSIGNEE(S):

Showa Sangyo Co., Ltd., Japan

SOURCE: Eur. Pat. Appl., 25 pp.

CODEN: EPXXDW

DOCUMENT TYPE:

Patent English

LANGUAGE:

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
EP 757098	A2	19970205	EP 1996-112114	19960726 <
EP 757098	A3	19970917		
R: DE, DK,	FR, GB	, IT		
JP 09037780	A2	19970210	JP 1995-213005	19950731 <
CA 2182059	AA	19970201	CA 1996-2182059	19960725 <
US 5827715	Α	19981027	US 1996-686647	19960726
US 5939308	Α	19990817	US 1998-131732	19980810
PRIORITY APPLN. INFO	. :		JP 1995-213005	19950731
			US 1996-686647	19960726

AB A heat-resistant maltose phosphorylase is provided from Bacillus sp. RK-1 and MK-1. It retains .gtoreq.80% activity after treatment in a buffer of pH 6.0 at 50-60.degree. for 15 min. Its optimum temp. was 55-70.degree., its pH optimum was 6.0-7.0 with stability retained at pH 5.5-8.0, the mol. wt. on gel filtration was 150-190 kDa and 75-90 kDa in SDS, and the isoelec. point 4.7-5.1.

The

heat-resistant maltose phosphorylase differs from known maltose phosphorylases in bacterial origin, optimum temp., and thermal stability. The enzyme or bacteria contg. the enzyme can be used for prepn. of .beta.-glucose-1-phosphoric or trehalose using. By carrying out enzymic reaction at high reaction temps. using this enzyme, it is possible to prep. .beta.-glucose-1-phosphoric acid or trehalose industrially advantageously, with lowering of contamination with various germs and shortening of reaction time.

ΡI	EP 757098 A2 1	9970205			
	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
ΡI	EP 757098	A2	19970205	EP 1996-112114	19960726 <
	EP 757098	A3	19970917		
	R: DE, DK,	FR, GB,	IT		
	JP 09037780	A2	19970210	JP 1995-213005	19950731 <
	CA 2182059	AA	19970201	CA 1996-2182059	19960725 <
	US 5827715	Α	19981027	US 1996-686647	19960726
	US 5939308	A	19990817	US 1998-131732	19980810
					- '11

AB A heat-resistant maltose phosphorylase is provided from Bacillus sp. RK-1 and MK-1. It retains .gtoreq.80% activity after treatment in a buffer of pH 6.0 at 50-60.degree. for 15 min. Its optimum temp. was 55-70.degree., its pH optimum was 6.0-7.0 with stability retained at pH 5.5-8.0, the mol. wt. on gel filtration was 150-190 kDa and 75-90 kDa in SDS, and the isoelec. point 4.7-5.1.

The

heat-resistant maltose phosphorylase differs from known maltose phosphorylases in bacterial origin, optimum temp., and thermal stability. The enzyme or bacteria contg. the enzyme can be used for prepn. of .beta.-glucose-1-phosphoric or trehalose using. By carrying out enzymic reaction at high reaction temps. using this enzyme, it is possible to prep. .beta.-glucose-1-phosphoric acid or trehalose industrially advantageously, with lowering of contamination with various germs and shortening of reaction time.

IT Bacillus stearothermophilus

```
(thermostable trehalose phosphorylase and; prepn.,
       thermostability, and synthetic use of heat-resistant maltose
       phosphorylase from Bacillus)
    9030-19-7P, Maltose phosphorylase 37205-59-7P, Trehalose
ΙT
    phosphorvlase
    RL: BAC (Biological activity or effector, except adverse); CAT (Catalyst
    use); PRP (Properties); PUR (Purification or recovery); BIOL (Biological
    study); PREP (Preparation); USES (Uses)
       (prepn., thermostability, and synthetic use of heat-resistant maltose
       phosphorylase from Bacillus)
L25 ANSWER 5 OF 9 HCAPLUS COPYRIGHT 2001 ACS
                       1996:476927 HCAPLUS
ACCESSION NUMBER:
                       125:108887
DOCUMENT NUMBER:
                       Thermostable trehalose phosphorylase
TITLE:
                       and its preparation with Bacillus stearothermophilus
                       Ishii, Keiko; Inoe, Yasushi; Tomita, Tetsuji
INVENTOR(S):
                       Showa Sangyo Co, Japan
PATENT ASSIGNEE(S):
                       Jpn. Kokai Tokkyo Koho, 10 pp.
SOURCE:
                       CODEN: JKXXAF
                       Patent
DOCUMENT TYPE:
                       Japanese
LANGUAGE:
FAMILY ACC. NUM. COUNT: 1
PATENT INFORMATION:
                                      APPLICATION NO. DATE
    PATENT NO. KIND DATE
    PATENT NO.
                                       -----
    JP 08131166 A2 19960528 JP 1994-295765 19941104 <--
    A novel trehalose phosphorylase is prepd. from the
    culture of Bacillus stearothermophilus strain SK-1 and characterized.
The
    enzyme exhibits a pH optimum 6.5-7.5, temp. optimum
    70-75.degree., pI 4.6-5.2, and mol. wt. 110-150 kDa by
    gel filtration. The enzyme remains >95% active after incubating at
     50-65.degree., pH 6.0 for 15 min.
    Thermostable trehalose phosphorylase and its
TΙ
    preparation with Bacillus stearothermophilus
    JP 08131166 A2 19960528 Heisei
PΙ
                                        APPLICATION NO. DATE
    PATENT NO.
                 KIND DATE
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                                        ______
    JP 08131166 A2 19960528 JP 1994-295765 19941104 <--
PΙ
    A novel trehalose phosphorylase is prepd. from the
AR
    culture of Bacillus stearothermophilus strain SK-1 and characterized.
The
     enzyme exhibits a pH optimum 6.5-7.5, temp. optimum
     70-75.degree., pI 4.6-5.2, and mol. wt. 110-150 kDa by
     gel filtration. The enzyme remains >95% active after incubating at
     50-65.degree., pH 6.0 for 15 min.
     trehalose phosphorylase prepn Bacillus
ST
     Bacillus stearothermophilus
TΤ
        (thermostable trehalose phosphorylase and prepn.
       with Bacillus stearothermophilus)
IT
     37205-59-7P, Trehalose phosphorylase
     RL: BPN (Biosynthetic preparation); PRP (Properties); BIOL (Biological
     study); PREP (Preparation)
        (thermostable trehalose phosphorylase and prepn.
       with Bacillus stearothermophilus)
L25 ANSWER 6 OF 9 HCAPLUS COPYRIGHT 2001 ACS
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ACCESSION NUMBER:

1995:906143 HCAPLUS

DOCUMENT NUMBER:

123:309235

TITLE:

Purification and characterization of trehalose

phosphorylase from Micrococcus varians

AUTHOR(S):

Kizawa, Hideki; Miyagawa, Ken-ichiro; Sugiyama,

Yoshio

SOURCE:

CORPORATE SOURCE:

Integrated Technology Laboratories, Takada Chemical

Industries, Ibaraki, 300-42, Japan

Biosci., Biotechnol., Biochem. (1995),

59(10), 1908-12

CODEN: BBBIEJ; ISSN: 0916-8451

DOCUMENT TYPE:

Journal

English LANGUAGE:

Trehalose phosphorylase (EC 2.4.1.64), which catalyzes AΒ the reversible reaction of phosphorolysis and synthesis of trehalose, was purified to homogeneity from a cell-free ext. of Micrococcus varians strain No. 39. The enzyme was shown to have a mol. wt. of 570,000 to 580,000 by gel filtration, and to have a subunit of mol. wt. of 105,000 by SDS-PAGE. The stoichiometry of the reaction between trehalose, Pi, glucose, and .beta.-glucose 1-phosphate was 1:1:1:1 (molar ratio). The enzyme had high specificity for trehalose,

glucose, and .beta.-glucose 1-phosphate. The Kms for trehalose, Pi, glucose, and .beta.-glucose 1-phosphate were 10, 3.1, 23, and 38 mM, resp. The kcats were 200 s-1 for trehalose phosphorolysis and 660 s-1 for trehalose synthesis. The enzyme was inhibited by validamycin A, validoxylamine A, 1-deoxynojirimycin, and Cu2+ during trehalose phosphorolysis, and by Cu2+, Zn2+, and Ni2+ during trehalose synthesis. Inhibition competitive against trehalose was noted with validamycin A, validoxylamine A, and 1-deoxynojirimycin. Initial velocity, product inhibition, and dead-end inhibition studies suggested that both trehalose phosphorolysis and trehalose synthesis proceeded through an ordered Bi Bi

- Purification and characterization of trehalose ΤI phosphorylase from Micrococcus varians
- Biosci., Biotechnol., Biochem. (1995), 59(10), 1908-12 SO CODEN: BBBIEJ; ISSN: 0916-8451
- Trehalose phosphorylase (EC 2.4.1.64), which catalyzes AB the reversible reaction of phosphorolysis and synthesis of trehalose, was purified to homogeneity from a cell-free ext. of Micrococcus varians strain No. 39. The enzyme was shown to have a mol. wt. of 570,000 to 580,000 by gel filtration, and to have a subunit of mol. wt. of 105,000 by SDS-PAGE. The stoichiometry of the reaction between trehalose, Pi, glucose, and .beta.-glucose 1-phosphate was 1:1:1:1 (molar ratio). The enzyme had high specificity for trehalose.

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- trehalose phosphorylase Micrococcus ST
- Kinetics, enzymic ΙT Michaelis constant Micrococcus varians

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(purifn. and characterization of trehalose
        phosphorylase from Micrococcus varians)
     7440-02-0, Nickel, biological studies
                                             7440-50-8, Copper, biological
Τጥ
              7440-66-6, Zinc, biological studies 19130-96-2,
                          37248-47-8, Validamycin A
                                                      38665-10-0,
     1-Deoxynojirimycin
    Validoxylamine A
    RL: BAC (Biological activity or effector, except adverse); BIOL
     (Biological study)
        (purifn. and characterization of trehalose
        phosphorylase from Micrococcus varians)
     37205-59-7P, Trehalose phosphorylase
     RL: BAC (Biological activity or effector, except adverse); BPR
(Biological
     process); PRP (Properties); PUR (Purification or recovery); BIOL
     (Biological study); PREP (Preparation); PROC (Process)
        (purifn. and characterization of trehalose
        phosphorylase from Micrococcus varians)
                                             99-20-7, Trehalose 14048-34-1,
     50-99-7, D-Glucose, biological studies
IT
                                 14265-44-2, Phosphate, biological studies
     .beta.-Glucose 1-phosphate
     RL: BPR (Biological process); BIOL (Biological study); PROC (Process)
        (purifn. and characterization of trehalose
        phosphorylase from Micrococcus varians)
L25 ANSWER 7 OF 9 HCAPLUS COPYRIGHT 2001 ACS
                         1995:538169 HCAPLUS
ACCESSION NUMBER:
DOCUMENT NUMBER:
                         123:7925
                         Production and application of maltose phosphorylase
TITLE:
                         and trehalose phosphorylase by a
                         strain of Plesiomonas
                         Yoshida, Masahiro; Nakamura, Nobuyuki; Horikoshi,
AUTHOR(S):
Koki
                         Res. Inst., Nihon Shokuhin Kako Co., Ltd., Fuji, 417,
CORPORATE SOURCE:
                         Japan
                         Oyo Toshitsu Kagaku (1995), 42(1), 19-25
SOURCE:
                         CODEN: OTKAE3; ISSN: 1340-3494
DOCUMENT TYPE:
                         Journal
                         English
LANGUAGE:
     A strain of Plesiomonas capable of producing intracellular thermostable
AΒ
     maltose phosphorylase (MP) and trehalose phosphorylase
     (TP), which were useful for the prodn. of trehalose from maltose, was
     isolated from mud of a Japanese seashore. The isolate (SH-35) grew well
     among the ranges of pH 6-9 and temp. 15-45.degree. with optima
     at pH 7.0 and 37.degree. by shaking cultivation, though the max.
     yield of these enzymes were obtained at pH 7.5 and 34.degree.
     for MP and pH 8.0 and 37.degree. for TP in a medium contg.
     maltose as a carbon source and mixt. of polypepton-S, yeast ext. and urea
     as nitrogen sources. Optimum pH and temp. for producing
     trehalose were pH 7-8 and 55-60.degree. in the presence of
     10-40% (wt./wt.) maltose and 5-50 mM inorg. phosphate,
     and about 60% (as dry basis) of maltose was converted into trehalose by
     the simultaneous action of both enzymes before and after extn. from cells
     under the best conditions. These microbial and enzymic characteristics
     are consistent with the industrial prodn. of trehalose from maltose.
     Production and application of maltose phosphorylase and trehalose
ΤI
     phosphorylase by a strain of Plesiomonas
     Oyo Toshitsu Kagaku (1995), 42(1), 19-25
SO
     CODEN: OTKAE3; ISSN: 1340-3494
     A strain of Plesiomonas capable of producing intracellular thermostable
AB
     maltose phosphorylase (MP) and trehalose phosphorylase
     (TP), which were useful for the prodn. of trehalose from maltose, was
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    the simultaneous action of both enzymes before and after extn. from cells
    under the best conditions. These microbial and enzymic characteristics
    are consistent with the industrial prodn. of trehalose from maltose.
    Plesiomonas maltose trehalose phosphorylase prodn
    application
ΙT
    Plesiomonas
        (prodn. and application of maltose phosphorylase and trehalose
       phosphorylase by strain of Plesiomonas)
     99-20-7P, Trehalose
TT
    RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (manuf. of, from maltose by maltose phosphorylase and trehalose
       phosphorylase from Plesiomonas)
     9030-19-7P, Maltose phosphorylase 37205-59-7P, Trehalose
ΙT
    phosphorylase
    RL: BAC (Biological activity or effector, except adverse); BMF
     (Bioindustrial manufacture); BIOL (Biological study); PREP (Preparation)
        (prodn. and application of maltose phosphorylase and trehalose
       phosphorylase by strain of Plesiomonas)
ΙT
     69-79-4, Maltose
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (trehalose manuf. from, by maltose phosphorylase and trehalose
       phosphorylase from Plesiomonas)
L25 ANSWER 8 OF 9 HCAPLUS COPYRIGHT 2001 ACS
ACCESSION NUMBER:
                         1975:27655 HCAPLUS
DOCUMENT NUMBER:
                         82:27655
                         Metabolism of trehalose in Euglena gracilis. Partial
TITLE:
                         purification and some properties of
phosphoglucomutase
                         acting on .beta.-glucose 1-phosphate
                         Belocopitow, Enrique; Marechal, Luis R.
AUTHOR(S):
                         Inst. Invest. Bioquim. "Fundacion Campomar", Buenos
CORPORATE SOURCE:
                         Aires, Argent.
                         Eur. J. Biochem. (1974), 46(3), 631-7
SOURCE:
                         CODEN: EJBCAI
DOCUMENT TYPE:
                         Journal
                         English
LANGUAGE:
     Phosphoglucomutase (I) for .beta.-D-glucose 1-phosphate (II) was purified
AB
     460-fold from cell-free exts. of E. gracilis var. bacillaris by treatment
     with protamine sulfate, gel filtration on Sephadex G-100, and chromatog.
     on a DEAE-cellulose column. I converted II reversibly into D-glucose
     6-phosphate (III). The optimum pH of the reaction was 7.0 and
     the equil. const., II/III was 0.035. I required .beta.-D-glucose
     1,6-diphosphate and a divalent cation such as Mg2+, Co2+, or Mn2+.
     Measurements on Sephadex G-100 gave an apparent mol. wt. of
     .apprx.27,000. I, together with a trehalose
     phosphorylase found in the same Euglena exts., would constitute a
     new catabolic pathway for trehalose.
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Eur. J. Biochem. (1974), 46(3), 631-7 SO CODEN: EJBCAI Phosphoglucomutase (I) for .beta.-D-glucose 1-phosphate (II) was purified AΒ 460-fold from cell-free exts. of E. gracilis var. bacillaris by treatment with protamine sulfate, gel filtration on Sephadex G-100, and chromatog. on a DEAE-cellulose column. I converted II reversibly into D-glucose 6-phosphate (III). The optimum pH of the reaction was 7.0 and the equil. const., II/III was 0.035. I required .beta.-D-glucose 1,6-diphosphate and a divalent cation such as Mg2+, Co2+, or Mn2+. Measurements on Sephadex G-100 gave an apparent mol. wt. of .apprx.27,000. I, together with a trehalose phosphorylase found in the same Euglena exts., would constitute a new catabolic pathway for trehalose. L25 ANSWER 9 OF 9 HCAPLUS COPYRIGHT 2001 ACS 1972:402254 HCAPLUS ACCESSION NUMBER: 77:2254 DOCUMENT NUMBER: Metabolism of trehalose in Euglena gracilis. TITLE: Partial purification and some properties of trehalose phosphorylase Marechal, Luis R.; Belocopitow, Enrique AUTHOR(S): Inst. Invest. Bioquim. Fund. Campomar, Buenos Aires, CORPORATE SOURCE: Argent. J. Biol. Chem. (1972), 247(10), 3223-8 SOURCE: CODEN: JBCHA3 DOCUMENT TYPE: Journal LANGUAGE: English Trehalose phosphorylase, an enzyme found in cell-free exts. of E. gracilis var. bacillaris, was purified 75-fold by treatment with protamine sulfate, centrifugation at 200,000 g, and chromatog. in a column of hydroxylapatite. This enzyme catalyzes the reversible phosphorolytic splitting of trehalose, yielding .beta.-glucose 1-phosphate and glucose as products. The optimum pH of the reaction was 7.0 for phosphorolysis and 6.3 for the synthesis of trehalose. The equil. const. changes with pH. It was 4.2 at pH 7.0 and 17 at pH 6.3. The enzyme is very unstable in the absence of inorg. phosphate, .alpha.- or .beta.-glucose 1-phosphate. Measurements in sucrose gradient gave a mol. wt. of 344,000. This enzyme together with a phosphoglucomutase for .beta.-glucose 1-phosphate found in the same Euglena exts. would constitute a new catabolic pathway for trehalose. Metabolism of trehalose in Euglena gracilis. I. Partial purification TΙ and some properties of trehalose phosphorylase J. Biol. Chem. (1972), 247(10), 3223-8 SO CODEN: JBCHA3 Trehalose phosphorylase, an enzyme found in cell-free AB exts. of E. gracilis var. bacillaris, was purified 75-fold by treatment with protamine sulfate, centrifugation at 200,000 g, and chromatog. in a column of hydroxylapatite. This enzyme catalyzes the reversible phosphorolytic splitting of trehalose, yielding .beta.-glucose 1-phosphate and glucose as products. The optimum pH of the reaction was 7.0

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sucrose gradient gave a mol. wt. of 344,000. This enzyme

together with a phosphoglucomutase for .beta.-glucose 1-phosphate found in the same Euglena exts. would constitute a new catabolic pathway for trehalose.

ST trehalose phosphorylase Euglena

IT Euglena gracilis (trehalose phosphorylase of)

IT 37205-59-7

RL: BOC (Biological occurrence); BIOL (Biological study); OCCU (Occurrence) (of Euglena gracilis);

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